RESEARCH ON SULPHUR-BACTERIA AND PHYSICAL-CHEMICAL PARAMETERS FROM SULPHUROUS WATERS OF JIBOU AREA

Szarita PASKUCZA¹, Rahela CARPA², Alexei REMIZOVSCHI², Teodor RUSU¹

¹University of Agricultural Sciences and Veterinary Medicine, Faculty of Agriculture 3-5 Manastur Way, Cluj-Napoca, Romania

²Babes-Bolyai University, Faculty of Biology and Geology, 1 M. Kogalniceanu Street, Cluj-Napoca, Romania

Corresponding author email: k_hella@yahoo.com

Abstract

Sulphur is the fourth major essential nutrient element, after nitrogen, phosphorus and potassium, involved in biogeochemical cycles in nature. Sulphur is essential in all organisms because it is an integral part of major metabolic compounds, such as amino acids (methionine and cysteine), and it is an active component of numerous co-factors and prosthetic groups (Fe–S clusters, sulpholipids, glucosinolates, vitamins, etc), which are involved in metabolic processes of organisms. Medical research appears to confirm the validity of the use of sulphurous spring waters as therapeutical and preventive remedies for a large variety of diseases. In order to evaluate the quality of sulphurous water, samples have been taken of the Sulphurous Springs in Jibou. The objectives of this study were to determine physical and chemical properties (pH, electrical conductivity, salinity, redox potential) of some sulphurous water samples of Jibou area, and also to determine the microbiota involved in S biogeochemical cycle. The physical analysis showed that the sulphurous water of Jibou falls within the very weakly acidic reaction class, and the salinity was 0.2% in all samples. The aerobic heterotrophic bacteria show a maximum value in the sample 4 (2630x10³ bacteria/ml). The microbial sulphurous community was detected in all water samples. This information should improve our understanding on adaptation mechanisms of bacteria involved in the sulphur cycle in correlation with physical-chemical parameters.

Key words: sulphurous water, Jibou, physical-chemical parameters, sulphate-reducing bacteria, sulphur-oxidation bacteria.

INTRODUCTION

Jibou is an important region for occurrence of therapeutic sulphurous waters in North-Western Romania. Natural sulphurous mineral waters from Jibou have a treatment potential mostly used in balneotherapy.

Sulphur is present in all organisms and occurs in organic compounds with catalytic, structural and regulatory functions, such as amino acids, proteins, enzymes, etc. On the other hand, the biological functions of inorganic sulphur compounds are limited. These compounds are needed in assimilation (prokaryotes, plants and fungi) and synthesis of organic compounds because they provide sulphur. Sulphate, sulphide and elemental sulphur are the most abundant sulphur substances, and provide evidence for the simultaneous occurrence of dissimilatory sulphate reduction and sulphide oxidation (at Eubacteria and Archaebacteria). Hydrogen sulphide is rarely present in natural waters, with the exception of special

sulphurous mineral springs. Hydrogen sulphide (H₂S) is the active component of sulphurous waters (Pokorna and Zabranska, 2015).

Balneotherapy treats many diseases, including immune-mediated skin diseases. Current medical research confirms the effect of sulphurous spring waters use, as preventive remedies and therapeutics for a large variety of diseases affecting the respiratory tract, skin, liver, intestine, gynaecological apparatus and osteoarticular system (Frosch, 2007). The different studies followed the effect of sulphurous medicinal waters in a murine dermatitis and on psoriatic patients. The role of hydrogen sulphide in releasing somatostatin during bathing treatment was also studied. Some results point out the anti-inflammatory role of sulphurous medicinal waters in psoriasis (Tsoureli-Nikita et al., 2002).

According to the World Health Organization (WHO, 2013) and US Environmental Protection Agency (EPA), water for human consumption should contain less than 250 ppm

of sulphates (Balintova et al., 2015; US EPA, 2017). When the sulphate concentrations for human consumption are compromised, the human health concerns would naturally arise (Backer, 2000; Fernando et al., 2018).

Other studies on sulphurous mineral waters revealed anti-oxidant properties (Albertini et al., 2007) and, under the same oral treatment, no pathological changes on biochemical oxidative enzymatic markers hematochrome were noticed in human healthy volunteers (Albertini et al., 2008). H₂S is as an endogenous interintracellular signaling molecule with multiple effects. Its most investigated roles are related to cardiovascular, nervous system inflammatory diseases (Matz et al., 2003). The effect of H₂S on various clinical pathology parameters has been so far restricted to acute hematologic and enzyme alteration studies.

Sulphur bacteria are prokaryotes that use reduced or oxidized sulphur compounds in their metabolism. Reduced sulphur compounds (sulphide and elemental sulphur) are involved as electron donors in photosynthesis of anoxygenic phototrophic sulphur bacteria, or as energy and electron sources by colourless sulphur-oxidizing bacteria. In opposition. oxidized sulphur compounds (sulphate), are involved as electron acceptors for the oxidation organic matter by sulphate-reducing bacteria. When the sulphur level in water is above the needs of plants and microbes the excess is settled down. All of it or only a part will further circulate through oxidation to sulphate or reduction to sulphide. Thus, it is important to know the bacteria involved in these reactions. Each group of sulphur bacteria needs available sulphur compounds in a specific redox status. Sulphur bacteria are a heterogeneous group with taxa belonging to different phylogenetic branches within Bacteria and Archaea.

Biological oxidation of sulphur is important in the natural sulphur cycle. The oxidation reactions are performed by sulphur-oxidizing bacteria from Alphaproteobacteria, Betaproteobacteria, Gammaproteobacteria, Epsilonproteobacteria, Chloroflexia and Chlorobia classes. The electrons resulted from sulphide oxidation are then used by aerobic chemotrophic colorless sulphur-oxidizing

bacteria (*Beggiatoa*, *Thiobacillus*, etc) in respiration and for autotrophic CO₂ reduction. The anaerobic phototrophic sulphur-oxidizing bacteria (green sulphur bacteria - *Chlorobium*, etc; purple sulphur bacteria - *Halochromatium*, *Thiocapsa*, etc) transfer electrons from sulphur or other donors for autotrophic CO₂ reduction, using light energy (Zang et al., 2018).

Sulphur reducers are able to reduce elemental sulphur, and are widespread among Bacteria and Archaea (Desulfurococcus, Clostridium, Methanococcus, Geobacter etc). They use elemental sulphur as an electron acceptor in oxidizing organic matter (formate, acetate, lactate, pyruvate, sugars etc). The sulphide produced can further pass to autotrophic sulphide-oxidizing bacteria (Thiobacillus, Halothiobacillus) which oxidize it back to elemental sulphur (Ghosh and Dam, 2009; Zang et al., 2018).

This paper aims to investigate the bacteria involved in sulphur cycle (aerobic and anaerobic bacteria) in relation to physical-chemical parameters in sulphurous waters from Jibou zone (Salaj County).

MATERIALS AND METHODS

Water samples. Different sulphurous mineral water samples were collected from several local springs from Jibou town area. For the sample collection ground-glass apparatus was used. All samples were kept in the lab at 4°C, waiting processing. The waiting time before processing didn't exceed 24 hours. The water samples for physical and chemical analysis were analysed according to SR EN ISO 5667-1:2007/AC, 2007 and SR EN ISO 5667-3, 2013, and they were also microbiologically studied according to SR EN ISO 19458, 2007. Physical and chemical analyses. A portable multiparameter was used for the physical and chemical analyses. The physical and chemical analysis consists in detection of some parameters as: pH, electrical conductivity (EC), salinity (%), redox potential (Eh) and dissolved oxygen (SR EN 27888, 1997; SR EN ISO 10523:2012).

Microbiological analyses. Bacteriological parameters for water samples were established, namely: the total number of the mesophilic heterotrophic bacteria (cfu = colony-forming

units), aerobic and anaerobic sulphur-reducing, sulphur-oxidising, sulphate-reducing bacteria. In order to determine these bacterial indicators from the water samples we used the methods according to Carpa et al., 2014 and different standards for water samples. The culture media used were: nutrient agar (for heterotophic bacteria) (SR EN ISO 6222, 2004); peptone water medium (for aerobic sulphur reducing bacteria) (Carpa et al., 2014); Oppenheimer and Gunkel culture media (for anaerobic sulphurreducing bacteria (Atlas, 2010); Starkey culture sulphate-reducing media (for Desulfovibrio) (Cusa, 1996); Postgate culture media (for sulphur-oxidizing bacteria (Atlas, 2010)).

Water samples were subjected to serial dilutions and the first three dilutions were used to inoculate all culture media. After inoculation, the samples were incubated for 7 days at a temperature of 25°C.

Growth examination. In order to assess the number of heterotrophic aerobic bacteria, the number of clusters on each Petri dish was counted and then the cfu/ml was computed. Bacteria involved in sulphur cycle were determined by multiple tubes method and data were interpreted according to the Alexander statistic table – 1965 (Carpa et al., 2014).

RESULTS AND DISCUSSIONS

The physical and chemical parameters were measured in all water samples with the portable multiparameter. The number of bacteria belonging to the following ecophysiological groups was assessed: aerobic mesophilic heterotrophs, aerobic sulphur-reducing bacteria. aerobic sulphur-oxidizing bacteria, anaerobic sulphur-reducing bacteria and anaerobic sulphate-reducing bacteria. The results indicated that the number of bacteria from each group varied depending on the sampling sites. Physical and chemical analyses. The actual acidity (pH) is very important because it influences physiological groups of bacteria. In the water samples the pH ranged between 6.73 and 6.88 (Table 1). These values belong the very weakly acidic reaction classes. The redox potential (Eh) is used in water assessments because measures the capacity of chemical elements to acquire electrons and by that be reduced. A high redox potential indicates a greater affinity of chemical species for electrons and a greater tendency to be reduced, therefore, an aerobic environment. Sulphur bacteria are critical mediators of redox transformations occurring within biogeochemical sulphur cycle of the biosphere. The values of redox potential in our water samples were very low. Values of electrical conductivity varied between 845 and 867 uS/cm. In all water samples was measured a salinity of 0.2%, which means that the sulphurous waters from Jibou area contain

Table 1. Physical and chemical parameters in water samples

salts.

Samples	pН	Eh (redox potential)	Electrical conductivity	Salinity (%)	Dissolved O ₂ (mg/l)
		(mV)	(µS/cm)		
1	6.73	-3	845	0.2	0.11
2	6.81	-5	858	0.2	0.250
3	6.84	-7	862	0.2	0.30
4	6.87	-9	860	0.2	0.10
5	6.88	-10	867	0.2	0.23

Microbiological analyses. The total number of mesophilic heterotroph bacteria was measured by the growth layers method. Based on the results, colony forming units (cfu/ml) were calculated.

Viable counts of aerobic and anaerobic sulphur-reducing, sulphur-oxidising, sulphate-reducing bacteria were assessed by most

probable number counts (MPN). The results are presented in Table 2, where one can observe that the predominant bacteria involved in the sulphur cycle are aerobic the sulphur-reducing bacteria, followed by sulphur-oxidizing ones. The number of anaerobic bacteria is quite low in all analysed water samples.

Samples	Bacteria						
	Heterotrophic bacteria	Sulphur oxidizing aerobic	Sulphur reducing aerobic	Sulphur reducing anaerobic	Sulphate reducing anaerobes		
	(cfu/ml)						
1	1146x10 ³	130	170	40	120	2.167	
2	1446x10 ³	380	440	68	91	2.835	
3	2093x10 ³	170	210	78	100	2.753	
4	2630x10 ³	170	310	82	94	2.806	
5	965v10 ³	140	260	61	82	2 640	

Table 2. Microbial communities in water samples

The aerobic heterotrophs constituted the largest eco-physiological group of bacteria and it is used as a global indicator of microbiological water quality (Figure 1).

The number of aerobic heterotrophs varied in those five water samples, exhibiting values

between 965x10³ cfu/ml and 2630x10³ cfu/ml. One can observe that the greatest abundance of aerobic heterotrophs was in water sample 4 and the smallest value was found in water sample 5.

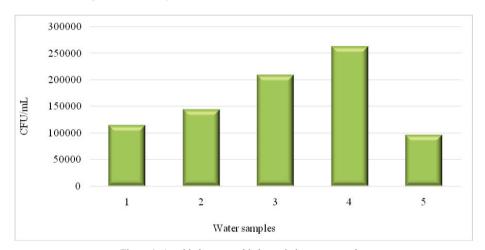


Figure 1. Aerobic heterotrophic bacteria in water samples

Bacteria in the Sulphur cycle removes the toxic effect of hydrogen sulphide and restore the sulphur to the biological use. They can be found on the bottom of standing waters and they are oxidizing the sulphur mineral compounds. Their energy source is hydrogen sulphide (H₂S), which they transform into sulphur (S), sulphurous acid (H₂SO₃) and sulphuric acid (H₂SO₄).

Sulphur-oxidizing bacteria are the main microorganisms that participate in the natural sulphur cycle. Sulphur-oxidizing bacteria are used for the continuous monitoring of toxicity and their use circumvents many of the obstacles associated with conventional assays (Luo et al., 2013). Sulphur-oxidizing bacteria, are chemoautotrophic bacteria that oxidize reduced sulphur compounds to sulphuric acid, in the presence of oxygen (O₂). Some of them, such as *Acidithiobacillus* genus, use elemental sulphur (S) particles as the electron donor (Sedky et al., 2013). Among aerobic bacteria involved in the sulphur cycle one can observe that aerobic sulphur-reducing bacteria are present in a slightly higher number compared to the number of aerobic sulphur-oxidizing bacteria (Figure 2).

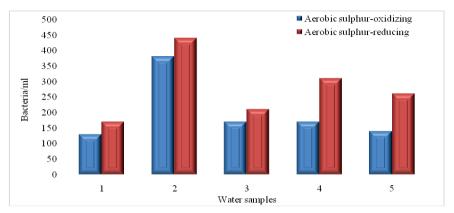


Figure 2. Aerobic sulphur bacteria (sulphur-oxidizing and sulphur-reducing) in water samples

Sulphur reducers can reduce elemental sulphur, and they can use elemental sulphur as the electron acceptor, to oxidize many organic matters (Ghosh and Dam. 2009).

Among the anaerobic bacteria, the anaerobic sulphur-reducing bacteria had the lowest count

in the water samples, compared to anaerobic sulphate-reducing bacteria and aerobic bacteria (figure 3). The lowest count was discovered to be for anaerobic sulphur-reducing bacteria in sample 1 (40 bacteria/ml).

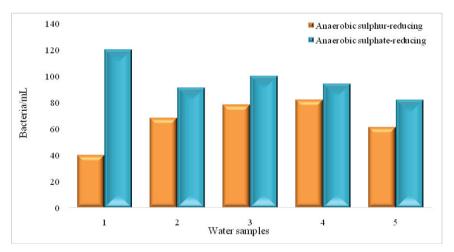


Figure 3. Anaerobic sulphur bacteria (sulphur-reducing and sulphate-reducing) in water samples

Results of the analyses indicated that the number of bacteria from each ecological group varied.

For a general characterisation of the sulphurous bacteria living in the water, we estimated the bacterial indicator of water quality (BIWQ), according to formula (Muntean, 1995-1996):

$$BIWQ = \frac{1}{n} \bullet \sum_{i=0}^{n} \log_{10} N$$

where:

BIWQ = the bacterial indicator of water quality;

n = the number of the physiological groups considered within the calculation;

N = the number of the bacteria belonging to each ecophysiological group.

For all samples, the BIWQ was very low and the calculated values ranged between 2.617 and 2.835. The highest BIWQ value was obtained in sample 2, collected from sulphurous waters from Jibou areas (Figure 4).

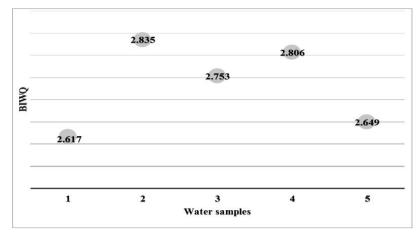


Figure 4. Bacterial indicators of water quality (BIWQ)

CONCLUSIONS

The physical analyses showed that the sulphurous water in Jibou area fits into the very weakly acidic reaction class. The values of redox potential in water samples were very low and these values affect the affinity of the nutrients and the presence of aerobic bacteria. Values of electrical conductivity varied between 845 and 867 μ S/cm. In all water samples a salinity of 0.2% was detected.

Numerical distribution of aerobic heterotrophic bacteria shows higher values in the sample 4 $(2630 \times 10^3 \text{ cfu/ml})$, followed by sample 3 $(2093 \times 10^3 \text{ cfu/ml})$ and the lowest value was found in sample 5 $(965 \times 10^3 \text{ cfu/ml})$.

The microbial sulphur community is not very abundant but suggests that all analysed groups were present in all water samples. Even if aerobic and anaerobic sulphur bacteria were present in the water samples, the aerobic sulphur bacteria were predominant.

The presence of all the five studied bacterial ecophysiological groups studied was noticed in all the water samples. Their number decreases in the following order: aerobic mesophilic heterotrophs $(965 \times 10^3 - 2630 \times 10^3 \text{ cfu/ml}) >$ aerobic sulphur-reducing bacteria (170 - 440 bacteria/ml) > aerobic sulphur-oxidizing bacteria (130 – 380 bacteria/ml) > anaerobic sulphate-reducing bacteria (82 bacteria/ml) > anaerobic sulphur-reducing bacteria (40 - 82 bacteria/ml). The bacterial indicator of water quality (BIWQ) had very low values, which varied between 2.617 and 2.835.

ACKNOWLEDGEMENTS

This work was supported by a grant of the Ministry of Research and Innovation, RDI Programe for Space Technology and Advanced Research – STAR, project number 699: HORUS – Software Toolbox for Pedological Monitoring of Transylvanian Area based on Sentinel-2 Data.

REFERENCES

Atlas R.M, 2010. Handbook of Microbiological Media, 4th edition, CRC Press, New York.

Albertini M.C., Teodori L., Accorsi A., Soukri A., Campanella L., Baldoni F., Dachà, M., 2008. Sulphurous mineral water oral therapy: Effects on erythrocyte metabolism Food and Chemical Toxicology 46: 3343–3350.

Albertini M.C., Dachà M., Teodori L., Conti M.E., 2007.
Drinking mineral waters: biochemical effects and health implications – the state-of-the-art. Int. J. Environ. Health 1: 153–169.

Backer L.C., 2000. Assessing the acute gastrointestinal effects of ingesting naturally occurring, high levels of sulfate in drinking water. Crit. Rev. Clin. Lab. Sci. 37 (4): 389–400.

Balintova M., Demcak S., Holub M., 2015. Sulphate removal from mine water-precipitation and bacterial sulphate reduction. In: Ali, M.A., Platko, P. (Eds.), Proceedings of the International Conference on Engineering Sciences and Technologies, Tatranská Štrba, High Tatras Mountains, Slovak Republic, pp. 239–244.

Carpa R., Dragan-Bularda M., Muntean V., 2014.
Microbiologie Generala-Lucrari Practice, Ed.

- Presa Univ. Clujeana, Cluj-Napoca [in Romanian].
- Cusa V., 1996. The methodological instruction for microbiological analysis of the aquatic sediments, vol. 4, Bucharest Research and Environmental.
- Fernando W.A.M., Ilankoon I.M.S.K, Syed T.H., Yellishetty M., 2018. Challenges and opportunities in the removal of sulphate ions in contaminated mine water: A review, Minerals Engineering, 117: 74–90.
- Frosch W.A., 2007. Taking the waters springs, wells, and spas. The FASEB Journal 21: 1948–1950.
- Ghosh W., Dam B., 2009. Biochemistry and molecular biology of lithotrophic sulfur oxidation by taxonomically and ecologically diverse bacteria and archaea, FEMS Microbiol. Rev. 33: 999– 1043
- Luo J., Tian G., Lin W., 2013. Enrichment, isolation and identification of sulfur-oxidizing bacteria from sulfide removing bioreactor, Journal of Environmental Sciences 25(7): 1393–1399.
- Matz H., Orion E., Wolf R., 2003. Balneotherapy in dermatology. Dermatologic Therapy, 16:132–40.
- Muntean V., 1995-1996. Bacterial indicator of mud quality, Contrib. Bot., Univ. Babes-Bolyai, Cluj-Napoca. p. 73.
- Pokorna D., Zabranska J., 2015. Sulfur-oxidizing bacteria in environmental technology, Biotechnology Advances 33: 1246–1259.
- Sedky H.A. Hassan S.H.A., Van Ginkel S.W., Sang-Eun Oh, 2013. Effect of organics and alkalinity on the sulfur oxidizing bacteria (SOB) biosensor, Chemosphere, 90: 965–970.
- Tsoureli-Nikita E., Menchini G., Ghersetich I., Hercogova J., 2002. Alternative treatment of

- psoriasis with balneotherapy using Leopoldine spa water, Journal of the European Academy of Dermatology and Venereology 16:260–2.
- Zhang Y., Zhang L., Li L., Chen G.H., Jiang F., 2018. A novel elemental sulfur reduction and sulfide oxidation integrated process for wastewater treatment and sulfur recycling. Chemical Engineering Journal, 342: 438–445.
- SR EN 27888, 1997. Calitatea apei. Determinarea conductivitatii electrice [in Romanian].
- SR EN ISO 6222, 2004. Calitatea apei. Numărarea microorganismelor de cultură. Numărarea coloniilor prin însămânțare în mediu de cultură nutritiv agar [in Romanian].
- SR EN ISO 5667-1;2007/AC, 2007. Calitatea apei. Prelevare. Partea 1: Ghid general pentru stabilirea programelor şi a tehnicilor de prelevare [in Romanian].
- SR EN ISO 19458, 2007. Calitatea apei. Prelevare pentru analiza microbiologica [in Romanian].
- SR EN ISO 10523, 2012. Calitatea apei. Determinarea pH-ului [in Romanian].
- SR EN ISO 5667-3, 2013. Calitatea apei. Prelevare. Partea 3: Conservarea si manipularea probelor de apa [in Romanian].
- US EPA, 2017. Secondary Drinking Water Standards:
 Guidance for Nuisance Chemicals.
 https://www.epa.gov/dwstandardsregulations/secondary-drinking-waterstandards guidance-nuisance-chemicals.
- WHO/UNICEF, 2013. Progress on Sanitation and Drinking-Water: 2013 Update. Geneva, Switzerland. Retrieved from http://apps.who.int/iris/bitstream/10665/81245/1/9 78924150 5390 eng.pdf.